Amendments to the Claim:

This listing of claims will replace all prior versions, and listings, of claims in the application:

Listing of Claims:

1 (previously presented). A method for maturation of conifer somatic embryos, comprising

an anti-auxin step, where an embryogenic cell mass is cultured with a culture medium comprising an anti-auxin.

2 (original). A method according to claim 1, further comprising a second step before the anti-auxin step, where the embryogenic cell mass is cultured with a culture medium.

3 (original). A method according to claim 1, further comprising a third step after the anti-auxin step where the embryogenic cell mass is cultured with a culture medium essentially free of anti-auxin.

4 (original). A method according to claim 1, whereby the anti-auxin step lasts from 2 days to 50 weeks.

5 (original). A method according to claim 2, whereby the second step before the anti-auxin step lasts from two days to 10 weeks.

6 (original). A method according to claim 3, whereby the third step after the anti-auxin step lasts from two days to 40 weeks.

7 (previously presented). A method according to claim 1, whereby the culture medium in at least one of the steps further comprises a maturation agent.

8 (original). A method according to claim 7, whereby the culture medium of all the steps further comprises at least one maturation agent.

9 (previously presented). A method according to claim 7, whereby the maturation agent is selected from the group comprising abscisic acid, silver nitrate, jasmonic acid, abscisyl alcohol, acetylenic aldehyde, dihydroacetylenic alcohol, phaseic acid, dihydrophaseic acid, 6'-hydroxymethyl abscisic acid, beta-

hydroxy abscisic acid, beta-methylglutaryl abscisic acid, beta-hydroxy-beta-methylglutarylhydroxy abscisic acid, 4'-desoxy abscisic acid, abscisic acid, beta-D-glucose ester, 2-2(2-p-chlorophenyl-transethyl-cyclopropane carboxylic acid.

10 (previously presented). A method according to claim 7, whereby the maturation agent is abscisic acid at a concentration of between 0.1 and 200 μM.

11 (cancelled).

12 (currently amended). A method according to claim 1, whereby the anti-auxin is selected from the group consisting of α-(1-naphtylmethyl-sulfide)-isobutyric acid, α-(1-naphtylmethyl-sulfide)-propionic acid, α-(2-naphytylmethyl-sulfide)-isobutyric acid, α-(2-naphtylmethyl-sulfide)-propionic acid, δ-(naphtylmethyl-selenide)-η-valeric acid, (-)-α-([[2,3,5]]2,4,5-trichlorophenoxy)-propionic acid, (-)-α-(2,-4-dichlorophenoxy)-propionic acid, (-)-α-(2-naphthoxy)-propionic acid, (+)-α-(1-naphthoxy)-propionic acid, (3-phenyl, 1,2,4-thiadiazol-5-yl)thioacetic acid (PTAA), β-naphthalene acetic acid (β-NAA), γ-phenylbutyric acid, 1-(naphthylmethyl-sulfide)-propionic acid, 1-naphthylmethyl-selenidacetic acid, 2-(naphthylmethyl-sulfide)-propionic acid, 2-(o-chlorophenoxy)-2-methylpropionic acid, 2,3,4,5,6-pentachlorophenoxyisobutyric acid, 2,3,5-tri-iodobenzoic acid (TIBA), ~~2,3,5-triiodobenzoic acid,~~ 2,4,5-trichlorophenoxyisobutyric acid, 2,4,6-trichlorophenoxyacetic acid (2,4,6-T), 2,4,6-trichlorophenoxyisobutyric acid, 2,4-dichloroanisole (2,4-DCA), 2,4-dichlorophenoxyisobutyric acid (2,4-DCIP), 2,4-dihlorophenylsulfoneacetic acid, 2,4-dichlorophenylsulfoxideacetic acid, 2,6-dichlorophenoxyacetic acid, 2-chlorophenoxyisobutyric acid, 2-naphtylmethyl-selenidacetic acid, 3-chlorophenoxyisobutyric acid, 3-indoleisobutyric acid, 3-nitro-4-flourobenzoid acid, 4-chlorophenoxyisobutyric acid, 5-methyltryptophan, 7-aza-indol, 9-hydroxyfluorene-9-carboxylic acid (HFCA), ferulic acid, flavonoids, indoleisobutyric acid, kaempferol, maleic hydrazide, naptalam (N-1-naphtylphthalamic acid), p-Chlorophenoxyisobutyric

acid (PCIB), p-coumaric acid, phenoxyacetic acid, phenoxyisobutyric acid, phenylpropionic acid, quercitin, and trans-cinnamic acid.

13 (cancelled).

14 (previously presented). A method according to claim 1, whereby the anti-auxin is PCIB at a concentration between 0.01 and 200 μM.

15 (original). A method according to claim 1, whereby the anti-auxin is PCIB at a concentration between 1 and 50 μM.

16 (currently amended). A method according to claim 1, whereby the conifer is a member of the ~~Pinaceae~~ *Pinaceae*.

17 (original). A method according to claim 1, whereby the conifer is selected from the genera *Pinus, Picea, Abies, Larix* and *Pseudotsuga*.

18 (previously presented). A method according to claim 1, whereby the conifer is an *Abies sp*.

19 (previously presented). A method according to claim 1, whereby the conifer is a *Picea sp*.

20 (previously presented). A method according to claim 1, whereby the conifer is an *Abies sp* and the anti-auxin is PCIB at a concentration between 1 and 100μM.

21 (original). A method according to claim 1, whereby the conifer is a *Picea sp* and the anti-auxin is PCIB at a concentration between 0.1 and 50 μM.

22 (original). A method according to claim 3, whereby the culture medium used during at least part of the third step after the anti-auxin step further comprises an auxin.

23-26 (cancelled).

27 (previously presented). A method according to claim 3, whereby the embryogenic cell mass is cultured with a culture medium comprising a carbohydrate source.

28 (previously presented). A method according to claim 27, where the embryogenic cell mass is cultured with a culture medium comprising sucrose, fructose, or glucose.

29-30 (cancelled).

31 (original). A method according to claim 27, whereby the culture medium has a content of between 1 and 100 g/L of metabolisable carbon sources.

32 (original). A method according to claim 27, whereby the further culturing is performed for a period of from 2 days to 10 weeks.

33-37 (cancelled).

38 (previously presented). A method according to claim 1 where the conifer is *Abies nordmannia*.

39. (previously presented). A method according to claim 1 where the conifer is *Picea abies* or *Pichea sitchenis*.